## Note

# Simple synthesis of [1-<sup>11</sup>C]acetate

D. Le Bars\*, M. Malleval, F. Bonnefoi and C. Tourvieille CERMEP - Imagerie du Vivant, 59 Bd PINEL, 69003 Lyon, France

#### Summary

 $[1-^{11}C]$ Acetate is prepared by carboxylation of a Grignard reagent, CH<sub>3</sub>MgBr, on a simple polyethylene loop with cyclotron-produced [<sup>11</sup>C]carbon dioxide, followed by hydrolysis and purification on solid-phase extraction cartridges. Copyright © 2006 John Wiley & Sons, Ltd.

Key Words: acetate; loop synthesis; solid-phase extraction; carbon 11

## Introduction

 $[1-{}^{11}C]$ Acetate is a well-known tracer entering the Krebs cycle, reflecting cell oxidative metabolism. Besides cardiology, a new interest is arising for  $[1-{}^{11}C]$ acetate in oncology mainly for the study of prostate and liver tumors.

 $[1-^{11}C]$ Acetate is prepared by carboxylation of a Grignard reagent, CH<sub>3</sub>MgBr or CH<sub>3</sub>MgCl, with cyclotron-produced  $[^{11}C]$ carbon dioxide, followed by hydrolysis and purification. The original synthesis<sup>1</sup> requires solvent extraction, a difficult step for automation; simplified methods have been reported to avoid phase separation, mostly making use of SPE (solid-phase extraction) techniques.<sup>2–5</sup> New techniques for <sup>11</sup>C-carboxylation, based on immobilization or containment of Grignard reagent on the inner surface of various tubing, have greatly improved the production of  $[1-^{11}C]$ acetate<sup>6</sup> and have been applied to other compounds such as WAY100635<sup>7</sup> and other syntheses (<sup>11</sup>C-methylations<sup>8</sup>).

For a new automated system, we aimed at a combination of these techniques to achieve 'the simplest system': loop method for Grignard carboxylation, based on results from Davenport *et al.*<sup>6</sup> and SPE techniques for purification, as reported for  $[1-^{11}C]$  acetate by Kruijer *et al.*<sup>3</sup> and Roeda *et al.*<sup>4</sup>

Copyright © 2006 John Wiley & Sons, Ltd.

<sup>\*</sup>Correspondence to: D. Le Bars, CERMEP - Imagerie du Vivant, 59 Blvd PINEL, 69003 Lyon, France. E-mail: lebars@univ-lyon1.fr

#### 264

# Experimental

#### Materials and methods

*Reagents*: Methyl magnesium bromide 3 M solution in diethyl ether, THF and other chemicals were obtained from Sigma Aldrich Fluka. Sodium bicarbonate for IV perfusion, NaHCO<sub>3</sub> 1.4% (0.16 mmol ml<sup>-1</sup>), is obtained from Aguettant.

Acetate carrier solution is  $1 \text{ mmol } \text{I}^{-1}$ . The loop was made from 1 m of 3.20 mm OD PE tubing (1.6 mm ID), washed before each run with 10 ml water, 10 ml THF and dried with 100 ml min<sup>-1</sup> nitrogen for 20–30 min. The solidphase cartridges were Alltech Maxi-clean IC-H, IC-OH and IC-Ag, 0.5 ml cartridges. H<sup>+</sup>/Ag column was obtained by mixing the contents of two of each SPE cartridges in a 2 ml syringe, equipped with rubber cap. Carbon 11 was obtained as [<sup>11</sup>C]carbon dioxide via the <sup>14</sup>N(p, $\alpha$ )<sup>11</sup>C reaction on N5.6 nitrogen with 0.5% N5.6 oxygen, with 18 MeV protons (IBA Cyclone 18/9 cyclotron). <sup>11</sup>CO<sub>2</sub> was concentrated in a stainless steel loop/cryogenic trap (liquid nitrogen).

Analytical HPLC was performed on a Beckman Gold system, with a Raytest GABI for radioactivity detection, or a Beckman RI detector on an Aminex HP-87H (organic acids)  $7.8 \times 300$  mm column eluted with 0.004 M H<sub>2</sub>SO<sub>4</sub> in water at 0.6 ml min<sup>-1</sup>, at room temperature. Radiochemical yields were determined with a Capintec CRC-120 dose calibrator.

# $[1-^{11}C]$ Acetate production

The loop is loaded under nitrogen through a 3-way stopcock with  $100 \,\mu$ l of CH<sub>3</sub>MgBr solution, freshly prepared from 1 ml of methyl magnesium bromide 3 M solution in diethyl ether diluted with 1 ml of distilled THF. A 5 ml min<sup>-1</sup> nitrogen flow is then established through the system, and cyclotron-produced <sup>11</sup>CO<sub>2</sub> is recovered from the target and released from the cryogenic trap through the loop less than 2 min after Grignard introduction.

Further, 2 ml of 1 mM carrier acetate solution is pushed through the loop and passed on the combination of SPE cartridges (mixed  $H^+/Ag$ ,  $OH^-$ ). Then 5 ml water and 10 ml air are used to rinse the loop and the cartridges, and then the anionic IC-OH SPE cartridge is washed with 10 ml of water before elution with 5 ml 0.9% NaCl solution in a vial containing 0.5 ml of 2 N HCl. A 200 ml min<sup>-1</sup> nitrogen flow (from an independent line) is established for 2 min for [<sup>11</sup>C]carbonate elimination.

The injectable [<sup>11</sup>C]acetate solution is obtained after neutralization with 6 ml NaHCO<sub>3</sub> 1.4%, filtration through 0.22  $\mu$ m millipore filter, and dilution with sterile isotonic saline (Figure 1).

#### **Results and discussion**

This simple setup produces  $[1^{-11}C]$  acetate in good yields (60–70% decay corrected to EOB). The amount of Grignard in this synthesis is similar to the amounts used by Kruijer *et al.* (0.3 mmol)<sup>3</sup> or Roeda *et al.* (0.1 mmol),<sup>4</sup> but higher than the amount (0.03 mmol) reported by Davenport *et al.* in their system based on a smaller teflon loop.<sup>6</sup> As shown by HPLC (Figure 2), by-products such as  $[^{11}C]$  acetone and  $[^{11}C]$ *t*-butanol are present in the waste fraction, with a minor amount of  $[1^{-11}C]$  acetate. SPE techniques afford an easy

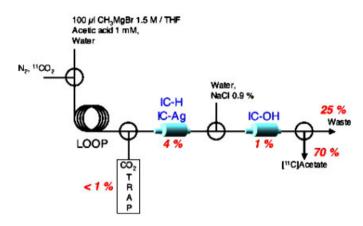


Figure 1. Synthesis apparatus and radioactivity distribution

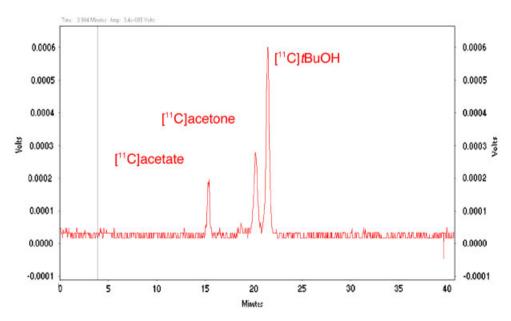


Figure 2. HPLC radiochromatogram of waste fraction

Copyright © 2006 John Wiley & Sons, Ltd.

J Label Compd Radiopharm 2006; 49: 263-267

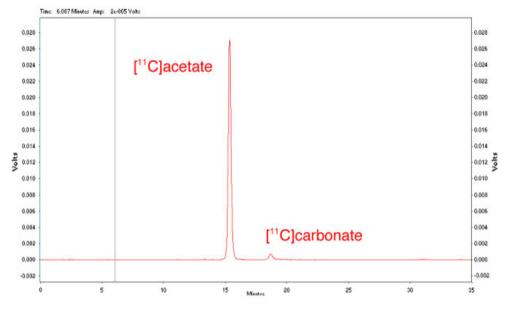


Figure 3. HPLC radiochromatogram of eluted fraction (NaCl 0.9%)

purification step, carrier acetate is added to ensure efficient and reproducible trapping and elution from the anion exchange column as shown by Kruijer *et al.*<sup>3</sup> In contrast to this similar purification process, we found that physiologic saline was sufficient to elute acetate from the anionic column. [<sup>11</sup>C]Carbonate amounted to 5–10% of radioactivity (Figure 3) and had to be removed with nitrogen flow in acidic solution. Analytical HPLC on Aminex HP 87-H reported the excellent chemical and radiochemical purities; HPLC analyses with refractive index detection did not show the unidentified contaminant reported by the Orsay group, with a different <sup>11</sup>CO<sub>2</sub> production system.<sup>4</sup> During the setup phase of the synthesis, preparative HPLC has been investigated: the carboxylation of the Grignard reagent can take place in a stainless-steel HPLC loop, and the reaction mixture can be directly injected on a C18 column to achieve HPLC purification (see Davenport *et al.*,<sup>6</sup> Kihlberg *et al.*<sup>9</sup> for HPLC use); this approach opens the possibility of acetate radiosynthesis in automated 'loop systems'.

#### Conclusion

Radiosynthesis has been automated with a programmable logic controller (PLC), a motor-driven syringe and electrovalves; the reaction is processed in less than 12 min and is afforded in a typical run of around 500 mCi, 18.5 GBq EOS-injectable [1-<sup>11</sup>C]acetate from an 18  $\mu$ Ah 36  $\mu$ A beam, in low specific radioactivity due to carrier-added purification, with excellent radiochemical and chemical purities.

#### Acknowledgements

Thanks to V.W. Pike, H. Tochon-Danguy and V. Tadino for fruitful discussions.

#### References

- 1. Pike VW, Horlock PL, Brown C, Clark JC. Appl Radiat Isotop 1984; 35: 623-627.
- 2. Iwata R, Ido T, Tada M. Appl Radiat Isotop 1995; 46: 117-121.
- 3. Kruijer P, Ter Linden T, Mooij R, Visser F, Herscheid DM. *Appl Radiat Isotop* 1995; **46**: 317–321.
- 4. Roeda D, Dolle F, Crouzel C. Appl Radiat Isotop 2002; 57: 857-860.
- 5. Moerlein SM, Gaehle GG, Welch MJ. Nucl Med Biol 2002; 29: 613-621.
- 6. Davenport RJ, Dowset K, Pike VW. Appl Radiat Isotop 1997; 48: 1117-1120.
- McCarron JA, Turton DR, Pike VW, Poole KG. J Label Compd Radiopharm 1996; 38: 941–953.
- 8. Wilson AA, Garcia A, Jin L, Houle S. Nucl Med Biol 2000; 27: 529-532.
- 9. Kihlberg T, Valind S, Langström B. Nucl Med Biol 1994; 21: 1067-1072.